

Lumbar Degenerative Disk Disease¹

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The sequelae of disk degeneration are among the leading causes of functional incapacity in both sexes and are a common source of chronic disability in the working years. Disk degeneration involves structural disruption and cell-mediated changes in composition. Mechanical, traumatic, nutritional, and genetic factors all may play a role in the cascade of disk degeneration, albeit to variable degree in different individuals. The presence of degenerative change is by no means an indicator of symptoms, and there is a very high prevalence in asymptomatic individuals. The etiology of pain as the symptom of degenerative disease is complex and appears to be a combination of mechanical deformation and the presence of inflammatory mediators. The role of imaging is to provide accurate morphologic information and influence therapeutic decision making. A necessary component, which connects these two purposes, is accurate natural history data. Understanding the relationship of etiologic factors, the morphologic alterations, which can be characterized with imaging, and the mechanisms of pain production and their interactions in the production of symptoms will require more accurate and reproducible stratification of patient cohorts.

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It has been stated that the term *degeneration*, as it is commonly applied to the intervertebral disk, covers such a wide variety of clinical, radiologic, and pathologic manifestations as to be really “only a symbol of our ignorance” (1). Despite this admonition, or perhaps because of it, we will attempt in this review to summarize current thoughts about the etiology, manifestations, and symptom production in lumbar degenerative disk disease and the role imaging currently plays in its identification and management. Given the gaps in our knowledge and the complexity of the subject, which will become readily apparent, we have chosen to exclude detailed treatment options from the discussion.

The sequelae of disk degeneration remain among the leading causes of functional incapacity in both sexes and are a common source of chronic disability in the working years. In accordance with its incidence, morbidity, and socioeconomic impact, degenerative disk disease has given and continues to give rise to extensive research efforts into its epidemiology, anatomy, biomechanics, biochemistry, and neuromechanisms (2).

Etiology

Traditionally, disk degeneration has been linked to mechanical loading. The

Essentials

- Mechanical, traumatic, nutritional, and genetic factors all play a role in the cascade of disk degeneration.
- Reliable and reproducible terminology is critical to meaningful description of morphologic abnormalities.
- The etiology of pain in degenerative disease is more complex than a simple mechanical explanation.
- The prognostic value of imaging is confounded by the high prevalence of morphologic changes in the asymptomatic population.
- In patients with uncomplicated low back pain or radiculopathy, MR imaging may not have an additive value over clinical assessment.

importance of mechanical factors has been emphasized by experiments on cadaver spines with both a severe single event and relentless loading (3–7). Failure of disks is more common in areas where there are the heaviest mechanical stresses, such as the lower lumbar region. It has been suggested that mechanical factors produce endplate damage, the antecedent to disk degeneration (8).

The disk is metabolically active, and the metabolism is dependent on diffusion of fluid either from the marrow of the vertebral bodies across the subchondral bone and cartilaginous endplate or through the annulus fibrosus from the surrounding blood vessels. Morphologic changes in the vertebral bone and cartilaginous endplate, which occur with advancing age or degeneration, can interfere with normal disk nutrition and further the degenerative process. This disruption of the normal endplate results in deformation when under loading. This allows nuclear material to pass through the endplate, reducing intradiscal pressure with subsequent bulging and loss of height and adding more stress to the surrounding annulus. Compressive damage to the vertebral body endplate alters the distribution of stresses in the adjacent disk. Continual cyclic loading makes these changes worse. Diminished blood flow in the endplate initiates tissue breakdown first in the endplate and then in the nucleus. These altered stress distributions adversely affect disk cell metabolism. These changes then alter the integrity of the proteoglycans and water concentration, reducing the number of viable cells with subsequent alteration in the movement of solutes into and out of the disk (9).

The importance of normal blood flow to the homeostatic nutritional process in the intervertebral disk complex has been suggested to explain the association of atherosclerosis and aortic calcification with increased disk degeneration and subjective low back pain (10). As degeneration progresses, structures of the disk become more disarranged and greater stresses are placed on the annulus and facet joints. As increased

forces are transmitted to the annulus, there may be fragmentation and fissuring. Disk degeneration involves structural disruption and cell-mediated changes in composition, but which occurs first is not clear. Biochemical factors can increase susceptibility to mechanical disruption, and this could adversely influence disk cell metabolism. Regardless of the initiating mechanism, these mechanisms would be interactive and additive, the end result being an altered functional ability of the disk to resist applied forces.

In addition to mechanical and nutritional causes, a genetic predisposition has been suggested by animal models that consistently develop degenerative disk disease at an early age, as well as by reports of familial osteoarthritis and lumbar canal stenosis in humans (11). In a study (12) of 115 male identical twin pairs, the effects of lifetime exposure to commonly suspected risk factors on disk degeneration, including job type, lifting, twisting, sitting, driving, exercise, trauma, and cigarette smoking, were investigated. The particular environmental factors studied, which have been among those most widely suspected of accelerating disk degeneration, had only modest effects. These small effects would help to explain the mixed results of previous studies. Considering the very minor effects the particular environmental factors studied had in determining disk degeneration, a strong genetic influence was suggested (12). While failing to find a strong association or clear-cut etiologic influence, the authors concluded that disk degeneration may be explained primarily by genetic influences and by unidentified

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Abbreviations:

ADC = apparent diffusion coefficient
 IL = interleukin
 MMP = matrix metalloproteinase
 SE = spin echo
 TNF = tumor necrosis factor

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factors, which may include complex unpredictable interactions.

In a cohort study (13) based on a Danish twin registry, substantial genetic influence on the susceptibility to degenerative disk disease and low back pain was shown. Shared environment was important until the age of 15 years, but as the twins grew older, the effect of a nonshared environment increased and nonadditive genetic effects became more evident. These findings suggest increasing genetic interaction (13).

Abnormalities of collagen are most often cited to support genetic influence in degenerative disk disease. Type II collagen is the most abundant collagen of cartilaginous tissues and is often referred to as the major collagen, forming heterotypic fibrils with the less abundant minor collagen types IX and XI. These fibrils provide the strength necessary to resist tensile forces. Disease that causes mutations in types II and XI collagen has been demonstrated in a number of chondrodystrophies. Multiple epiphyseal dysplasia is associated with a disease causing mutation in collagen IX, an important structural component of the annulus fibrosus, nucleus pulposus, and hyaline cartilage of the endplates. In a study looking for additional disease-producing mutations, researchers closely examined the genes coding the three chains of type IX collagen. While no changes typical of disease producing mutations were identified, two amino acid substitutions were identified that are substantially more prevalent in patients with lumbar degenerative disk disease than in normal controls. Both of these involve a tryptophan substitution (Trp2 and Trp3). In the case of the Trp3 allele, it is a genetic factor associated with a threefold increase in the risk of symptomatic degenerative disk disease (14–16). While these Trp alleles together occurred in 16% of the Finnish patients with lumbar disk disease, it has been pointed out that other loci are almost certainly involved. A prime candidate is one that encodes aggrecan, an abundant proteoglycan, in cartilage whose extensive hydration contributes to resistance to tissue deformation. Knock-out mice for aggrecan have a high

prevalence of disk herniation and degeneration (15,17).

Several additional studies (18–21) suggest that not just the process of degenerative disk disease but perhaps even its sequelae, including disk herniation, low back pain, and radiculopathy, are strongly influenced by genetic factors. Studies suggest that low back pain is also associated with polymorphism in the interleukin (IL) 1 locus (22). This is of interest in that cytokines such as tumor necrosis factor α (TNF- α), IL-1, and IL-6 are important inflammatory mediators, as will be discussed later.

Clearly there are many interactive factors at play. Mechanical, traumatic, nutritional, and genetic factors all may play a role in the cascade of disk degeneration, albeit to variable degrees in different individuals. Whatever the etiology, by the age of 50 years, 85%–95% of adults show evidence of degenerative disk disease at autopsy (23).

Morphologic Alterations and Imaging

Terminology

No less a problem than understanding etiology is agreeing on terminology that is reliable and reproducible to describe the morphologic alterations produced by the degenerative process (24–27). For the purposes of this review, we have used the terminology described by Milette (28). The term *degeneration* includes any or all of the following: real or apparent desiccation, fibrosis, narrowing of the disk space, diffuse bulging of the annulus beyond the disk space, extensive fissuring (ie, numerous annular tears) and mucinous degeneration of the annulus, defects and sclerosis of the endplates, and osteophytes at the vertebral apophyses. At magnetic resonance (MR) imaging, these changes are manifested by disk space narrowing, T2-weighted signal intensity loss from the intervertebral disk, presence of fissures, fluid, vacuum changes and calcification within the intervertebral disk, ligamentous signal changes, marrow signal changes, osteophytosis, disk herniation, malalignment, and stenosis. While there is confusion in the differen-

tiation of changes of the pathologic degenerative process in the disk from those of normal aging, we will use the term *degenerative* to include all such changes (29–31).

Conventional theory would imply that degeneration and aging are very similar processes, albeit occurring at different rates (32). Resnick and Niwayama (32) emphasized the differentiating features of two degenerative processes involving the intervertebral disk, which had been previously described by Schmorl and Junghans (33). These include “spondylosis deformans,” which affects essentially the annulus fibrosus and adjacent apophyses, and “intervertebral osteochondrosis,” which affects mainly the nucleus pulposus and the vertebral body endplates but also includes extensive fissuring (numerous tears) of the annulus fibrosus. Scientific studies suggest that spondylosis deformans is the consequence of normal aging, whereas intervertebral osteochondrosis, sometimes also called deteriorated disk, results from a clearly pathologic, though not necessarily symptomatic, process (33–38). Anterior and lateral marginal vertebral body osteophytes have been found in 100% of skeletons of individuals over 40 years of age, and therefore are consequences of normal aging, whereas posterior osteophytes have been found in only a minority of skeletons of individuals over 80 years, and therefore are not inevitable consequences of aging (34). Endplate erosions with osteosclerosis and chronic reactive bone marrow changes also appear to be pathologic.

Anatomic Considerations

The intervertebral joint is a three-joint complex consisting of the endplate-disk-endplate joint of the anterior column and the two facet joints of the posterior column supported by ligaments and muscle groups. Understanding the interrelationship of these elements has become more critical as surgical intervention, much like joint surgery in the past, is transitioning from fusion to joint replacement.

The intervertebral disk and the diarthrodial joints (zygapophyseal joint or

facet joints) interactively degenerate, causing altered stresses on the integrity and mechanical properties of the spinal ligaments, which results in degeneration of the spinal unit as a whole (39,40). The manner of degeneration of the various components of the spine is mediated and manifested by the specific structure involved. The cartilaginous, synovial, and fibrous structures each degenerate in a specific manner, which is associated with characteristic imaging and pathologic aberrations.

Degenerative Disk Changes

The major cartilaginous joint (amphiarthrosis) of the vertebral column is the intervertebral disk. Each disk consists of an inner portion, the nucleus pulposus, surrounded by a peripheral por-

tion, the annulus fibrosus. The nucleus pulposus is eccentrically located and more closely related to the posterior surface of the intervertebral disk. With degeneration and aging, type II collagen increases outwardly in the annulus and there is a greater water loss from the nucleus pulposus than from the annulus. This results in a loss of the hydrostatic properties of the disk, with an overall reduction of hydration in both areas to about 70%. In addition to water and collagen, the other important biochemical constituents of the intervertebral disk are the proteoglycans. The individual chemical structures of the proteoglycans are not changed with degeneration, but their relative composition is. The ratio of keratin sulfate to chondroitin sulfate increases, and there is a

diminished association with collagen that may reduce the tensile strength of the disk. The decrease in water-binding capacity of the nucleus pulposus is thought to be related to the decreased molecular weight of its nuclear proteoglycan complexes (aggregates). The disk becomes progressively more fibrous and disorganized, with the end stage represented by amorphous fibrocartilage and no clear distinction between nucleus and annulus (41–43). On T2-weighted images, the central disk signal intensity is usually markedly decreased and at distinct variance to that seen in unaffected disks of the same individual. Work with T2-weighted spin-echo (SE) sequences (44) suggests that MR is capable of depicting changes in the nucleus pulposus and annulus fibrosus relative to degeneration and aging based on a loss of signal intensity.

In work with cadaver spines of various ages, absolute T2 measurements correlated more closely with glycoaminoglycan concentration than absolute water content. Thus, the signal intensity may not be related to the total amount of water but rather the state of water. At present, the role that specific biochemical changes (proteoglycan ratios, aggregation of complexes) play in the altered signal intensity is not well understood. Given that the T2 signal intensity in the disk appears to track the concentration and regions of high glycoaminoglycan percentages more than absolute water content, it seems likely that the health and status of the proteoglycans are major determinates of signal intensity (45).

It has been proposed that annular disruption is the critical factor in degeneration and, when a radial tear develops in the annulus, there is shrinkage with disorganization of the fibrous cartilage of the nucleus pulposus and replacement of the disk by dense fibrous tissue with cystic spaces (31,46–49). Annular tears, also properly called annular fissures, are separations between annular fibers, avulsion of fibers from their vertebral body insertions, or breaks through fibers that extend radially, transversely, or concentrically and involve one or many layers of the annular

Figure 1

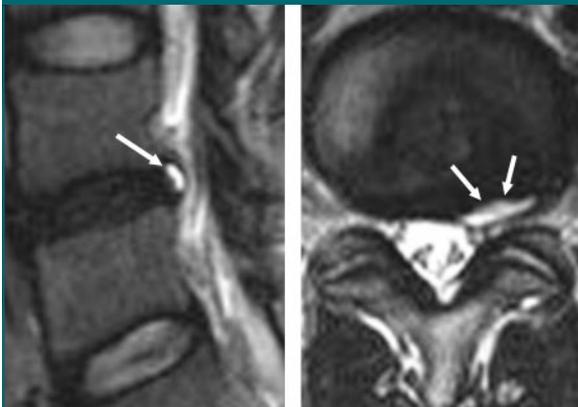


Figure 1: Annular tear. (a) Left parasagittal and (b) transverse T2-weighted fast SE (repetition time msec/echo time msec, 4000/120) MR images through L4-5 disk space. Note high signal intensity in the outer annulus and/or longitudinal ligament complex in a left parasagittal location, which represents area of annular disruption (arrows).

Figure 2

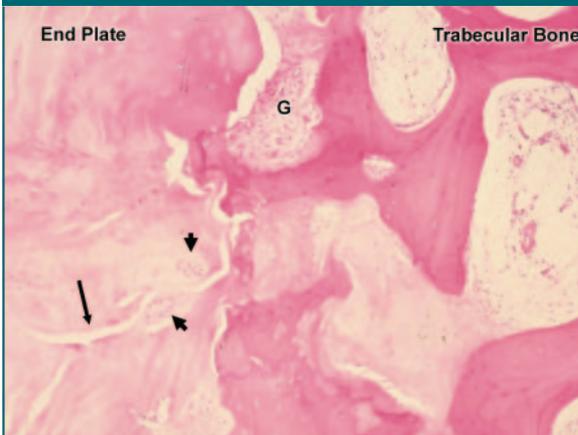


Figure 2: Histologic slide of degenerated intervertebral disk. Endplate shows areas of cracks, fissures, and pale staining (long arrow). Chondrocytes (short arrows) and granulation tissue (G), characteristic of regeneration and degeneration, are noted. Adjacent marrow space shows increased lipid elements and the trabeculae appear thickened.

lamellae. The term *tear* or *fissure* describes the spectrum of such lesions and does not imply that the lesion is consequent to trauma (Fig 1). Although it has certainly been verified that annular disruption is a sequela of degeneration and certainly is often associated with it, its role as the causal agent of disk degeneration has certainly not been proved. MR is the most accurate anatomic method for assessing intervertebral disk disease. The signal intensity characteristics of the disk on T2-weighted images reflect changes caused by aging or degeneration. A classification scheme for lumbar intervertebral disk degeneration has been proposed that has reasonable intra- and interobserver agreement (50). To date, however, there has been no correlation between MR disk changes and patient's symptoms.

With loss of water and proteoglycans, the nucleus pulposus is desiccated and friable with yellow-brown discoloration. Its onion-skin appearance begins to unravel, and cracks, clefts, or crevices appear within the nucleus and extend into the annulus fibrosus. Fissuring, chondrocyte generation, and granulation tissue formation may be noted within the endplate, annulus fibrosus, and nucleus pulposus of degenerative disks, indicating attempts at healing (49) (Fig 2). Radiolucent collections (vacuum disk phenomena) representing gas, principally nitrogen, occur at sites of negative pressure produced by the abnormal spaces (51). The vacuum phenomenon within a degenerated disk is represented on SE images as areas of signal void (52). Whereas the presence of gas within the disk is usually suggestive of degenerative disease, spinal infection may (rarely) be accompanied by intradiscal or intraosseous gas (53).

As intervertebral osteochondrosis progresses, there may be calcification of the disk. Calcification has usually been described on MR images as a region of decreased or absent signal intensity. The loss of signal is attributed to a low mobile proton density, as well as, in the case of gradient-echo imaging, to its sensitivity to the heterogeneous magnetic susceptibility found in calcified tissue. There is, however, variability in

Figure 3

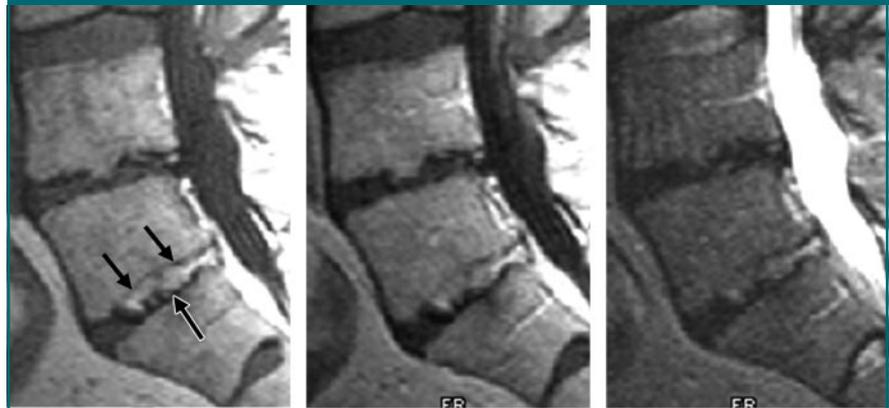


Figure 3: Calcified intravertebral disk. Sagittal midline (a) T1-weighted (450/12), (b) contrast-enhanced T1-weighted (450/12), and (c) T2-weighted (4000/120) SE images of lumbar spine. High signal intensity (arrows) is identified within L5-S1 disk space on a and b.

signal intensity of calcium at various sequences, and the type and concentration of calcification are important factors. Hyperintense disks on T1-weighted MR images may be secondary to calcification (Fig 3) (54). For concentrations of calcium particulate up to 30% by weight, the signal intensity on standard T1-weighted images increased but then subsequently decreased (55,56). These data likely reflect particulate calcium reducing T1 relaxation times by a surface-relaxation mechanism. Hyperintensities that are affected by fat-suppression techniques have also been noted within intervertebral disks and are thought to be related to ossification with lipid marrow formation in severely degenerated or fused disks.

Degenerative Marrow Changes

The relationship among the vertebral body, endplate, annulus, and disk has been studied (57–59) by using both degenerated and chymopapain-treated disks as models. Signal intensity changes in vertebral body marrow (Fig 4) adjacent to the endplates of degenerated disks are a common observation on MR images and appear to take three main forms.

Type I changes demonstrate decreased signal intensity on T1-weighted images and increased signal intensity on T2-weighted images and have been

Figure 4

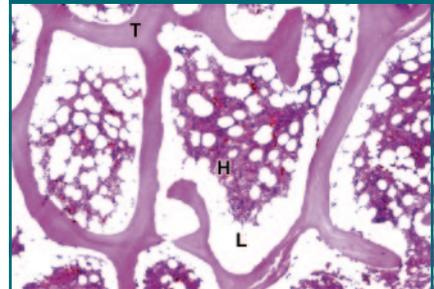


Figure 4: Normal marrow. Histologic slide of normal hematopoietic (H), lipid (L), and bone trabecular (T) elements.

identified in approximately 4% of patients scanned for lumbar disease (Fig 5), approximately 8% of patients after discectomy (60,61), and in 40%–50% of chymopapain-treated disks, which may be viewed as a model of acute disk degeneration (58). Histopathologic sections of disks with type I changes show disruption and fissuring of the endplate and vascularized fibrous tissues within the adjacent marrow, prolonging T1 and T2. Enhancement of type I vertebral body marrow changes is seen with administration of gadopentetate dimeglumine that at times extends to involve the disk itself and is presumably related to the vascularized fibrous tissue within the adjacent marrow (Fig 6).

Type II changes are represented by increased signal intensity on T1-weighted images and isointense or slightly hyperintense signal on T2-weighted images and have been identified in approximately 16% of patients at MR imaging (Fig 7). Disks with type II changes also show evidence of endplate disruption, with yellow (lipid) marrow replacement in the adjacent vertebral body resulting in a shorter T1 (Fig 8).

Type III changes are represented by a decreased signal intensity on both T1- and T2-weighted images and correlate with extensive bony sclerosis on plain

radiographs. The lack of signal in the type III change no doubt reflects the relative absence of marrow in areas of advanced sclerosis (Fig 9). Unlike type III, types I and II changes show no definite correlation with sclerosis at radiography (59). This is not surprising when one considers the histology; the sclerosis seen on plain radiographs is a reflection of dense woven bone within the vertebral body, whereas the MR changes are more a reflection of the intervening marrow elements.

Similar marrow changes have also been noted in the pedicles. While origi-

nally described as being associated with spondylolysis, they have also been noted in patients with degenerative facet disease and pedicle fractures. Again, the changes are probably a reflection of abnormal stresses, be they loading or motion (62,63).

Degenerative Facet and Ligamentous Changes

The superior articulating process of one vertebra is separated from the inferior articulating process of the vertebra above by a synovium-lined articulation, the zygoapophyseal joint. Like all diarthrodial synovium lined joints, the lumbar facet joints are predisposed to arthropathy with alterations of the articular cartilage. With disk degeneration and loss of disk space height, there are increased stresses on the facet joints with craniocaudal sub-

Figure 5

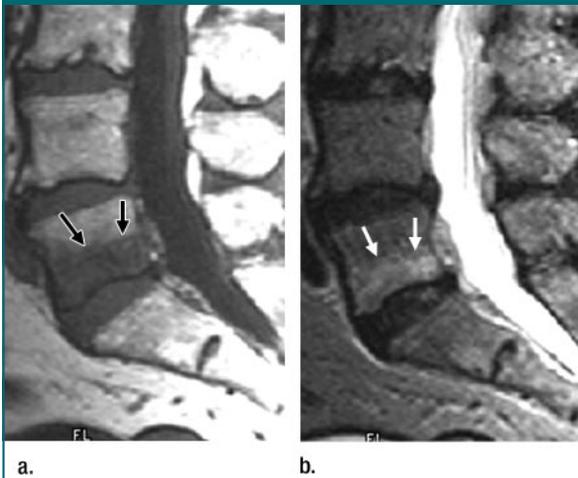


Figure 5: Degenerative type I marrow change. **(a)** Sagittal midline T1-weighted SE (450/12) image demonstrates decreased signal intensity of marrow space adjacent to L5-S1 disk (arrows). **(b)** T2-weighted SE (4000/120) image in the same region (arrows) shows increased signal intensity.

Figure 6



Figure 6: Histologic slide of type I degenerative marrow changes. Fibrovascular tissue (FV) has replaced normal marrow elements between thickened bone trabeculae (T).

Figure 7

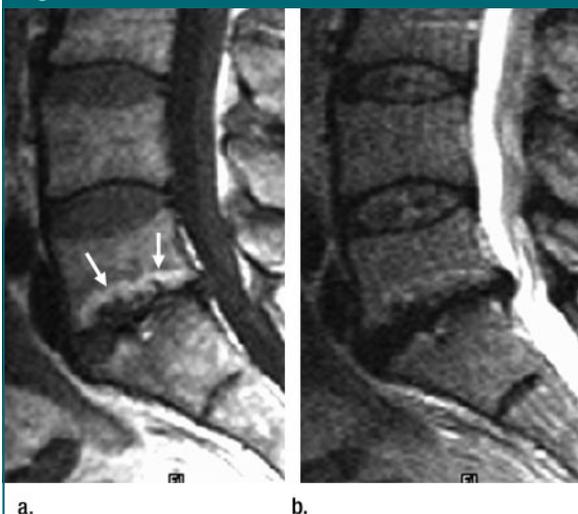


Figure 7: Degenerative type II vertebral body marrow changes. **(a)** Sagittal midline T1-weighted SE (450/12) image demonstrates degenerative changes of L5-S1 disk space and high signal intensity (arrows) in adjacent vertebral body marrow. **(b)** T2-weighted SE (4000/120) image shows that marrow signal intensity adjacent to degenerated L5-S1 disk is now only slightly hyperintense relative to more normal marrow.

Figure 8

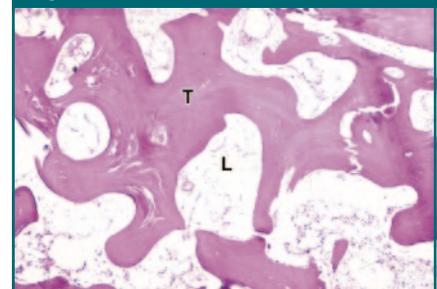


Figure 8: Histologic slide of degenerative type II marrow changes shows increased lipid content of the marrow space (L). Note also thickened woven bone trabeculae (T).

luxation resulting in arthrosis and osteophytosis. The superior articular facet is usually more substantially involved. Facet arthrosis can result in narrowing of the central canal, lateral recesses, and foramina and is an important component of lumbar stenosis (Fig 10). However, it has been proposed that facet arthrosis may occur independently and be a source of symptoms on its own (64,65). Syno-

vial villi may become entrapped within the joint with resulting joint effusions. The mechanism of pain may be related to nerve root compression from degenerative changes of the facets or by direct irritation of pain fibers from the innervated synovial linings and joint capsule (65). Osteophytosis and herniation of synovium through the facet joint capsule may result in synovial cysts, although the etiology of these

facet joint cysts is unclear (Fig 11). There is a more straightforward relationship of synovial cysts with osteoarthritis and the instability of the facet joints than degeneration of the intervertebral disk alone. In a review of patients with degenerative facet disease, synovial cysts occurred at anterior or intraspinal location in 2.3% of cases and posterior or extraspinal location in 7.3% (66).

The important ligaments of the spine include the anterior longitudinal ligament, the posterior longitudinal ligament, the paired sets of ligamenta flava (connecting the laminae of adjacent vertebrae), intertransverse ligaments (extending between transverse processes), and the unpaired supraspinous ligament (along the tips of the spinous processes). As these ligaments normally provide stability, any alteration in the vertebral articulations can lead to ligamentous laxity with subsequent deterioration. Loss of elastic tissue, calcification and ossification, and bone proliferation at sites of ligamentous attachment to bone are recognized manifestation of such degeneration.

Excessive lordosis or extensive disk space loss in the lumbar spine leads to close approximation and contact of spinous processes and to degeneration of

Figure 9

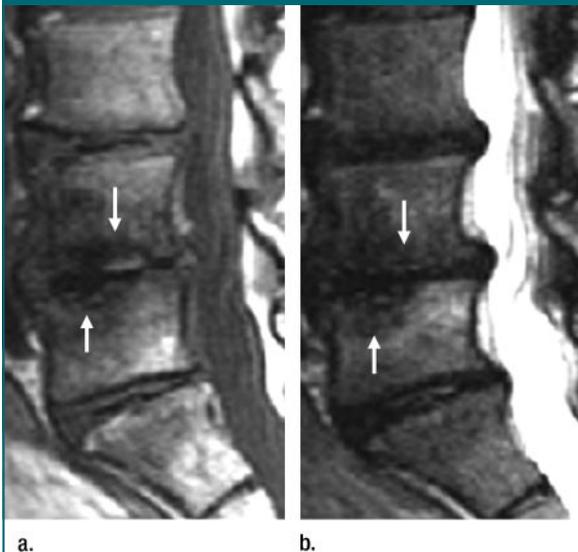


Figure 9: Degenerative type III vertebral body marrow changes. Sagittal midline (a) T1-weighted (450/12) and (b) fast SE T2-weighted (4000/120) images demonstrate markedly decreased signal intensity of adjacent marrow spaces at L4-5 in the presence of severe degenerative disk disease (arrows).

Figure 10



Figure 10: Spinal stenosis. Sagittal (a) T1-weighted SE (450/15) and (b) T2-weighted fast SE (4000/120) images through lower lumbar spine. There is grade I degenerative spondylolisthesis at L4-5, thickening of the ligaments posteriorly, and severe stenosis of central canal (arrows). Transverse (c) T1-weighted (450/15) and (d) T2-weighted fast SE (4000/120) images through L4-5 disk space show severe bilateral degenerative facet changes with distraction and fluid in the left joint (arrows). There is severe central canal stenosis and thickening of posterior ligaments.

intervening ligaments (67,68). Histologically, granulomatous reaction and perivascular cellular infiltration characterize the condition (Fig 12).

Morphologic and Functional Sequellae

Common, potential complications of degenerative disk disease include alignment abnormalities, intervertebral disk

displacement, and spinal stenosis. Various types of alignment abnormalities can exist alone or in combination, but the two most frequent are segmental instability and spondylolisthesis.

Instability

Segmental instability can result from degenerative changes involving the in-

tervertebral disk, vertebral bodies, and facet joints that impair the usual pattern of spinal movement, producing motion that is irregular, excessive, or restricted. It can be translational or angular.

Spondylolisthesis results when one vertebral body becomes displaced relative to the next most inferior vertebral body. The most common types include degenerative, isthmic, iatrogenic, and traumatic. Degenerative spondylolisthesis is seen usually with an intact pars interarticularis, is related primarily to degenerative changes of the apophyseal joints, and is most common at the L4-5 vertebral level (Fig 10). Predisilection for degenerative spondylolisthesis at that level is thought to be related to the more sagittal orientation of the facet joints, which makes them increasingly prone to anterior displacement. Degenerative disk disease may predispose to or exacerbate this condition secondary to narrowing of the disk space, which can pro-

Figure 11

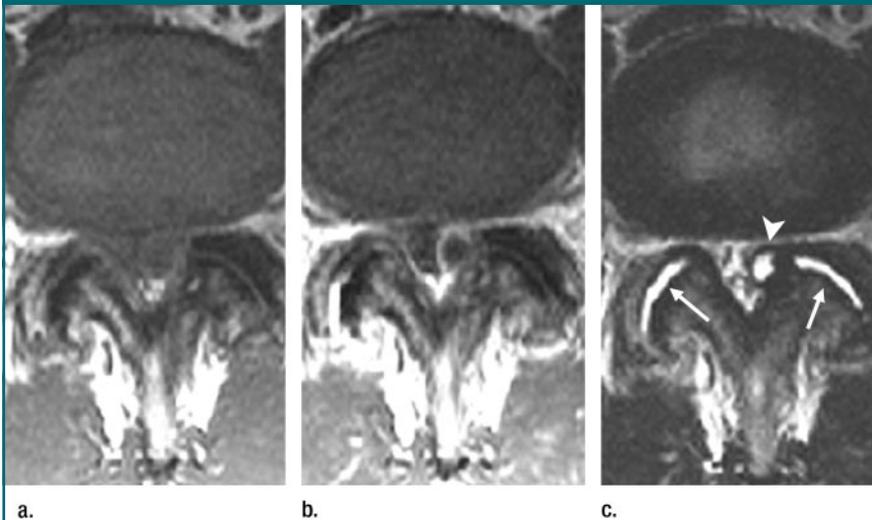


Figure 11: Degenerative synovial cyst. Transverse (a) T1-weighted SE (450/15), (b) T1-weighted contrast-enhanced SE (450/15), and (c) T2-weighted fast SE (4000/120) images through L4-5 disk space. Note severe bilateral degenerative facet changes and degenerative synovial cyst projecting medially from the left facet, causing central canal stenosis (arrowhead). There is distraction and fluid within degenerated facet joints (arrows).

Figure 13

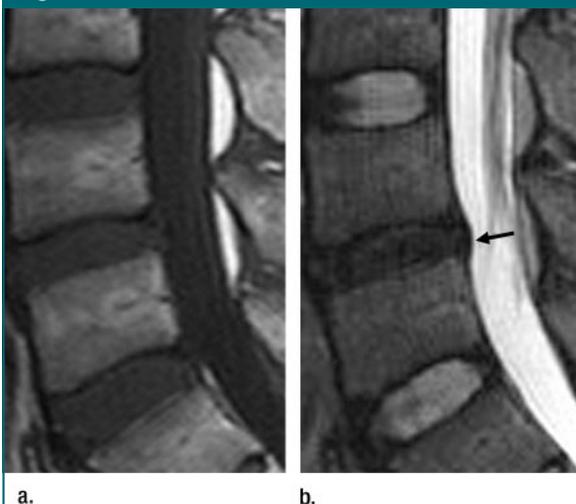


Figure 13: Degenerated L4-5 disk. Sagittal midline (a) T1-weighted SE (450/15) and (b) T2-weighted fast SE (4000/120) images show mild reduction in L4-5 disk space and loss of signal intensity on b. There is mild convex posterior bulging of the intervertebral disk at this level (arrow). Note normal signal intensity and morphology of L3-4 and L5-S1 disks.

Figure 12

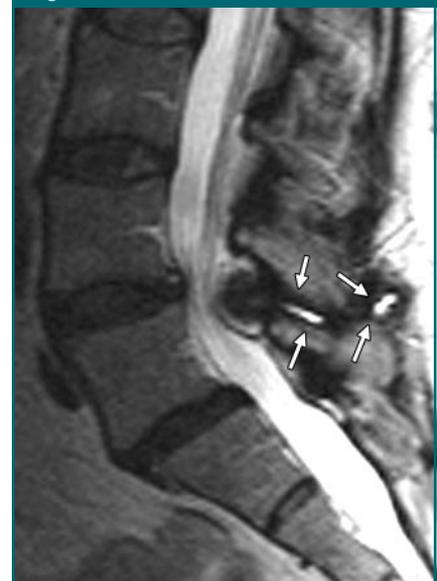


Figure 12: Severe degenerated posterior ligaments. Sagittal midline T2-weighted SE (4000/120) image shows mild grade I degenerative spondylolisthesis at L4-5. There is close approximation of spinous processes of L4 and L5 and high-signal-intensity degenerative changes in the region of intraspinal ligaments (arrows) (Bastrop disease).

duce subsequent malalignment of the articular processes and lead to rostral-caudal subluxation.

Herniation

Herniation refers to localized displacement of nucleus, cartilage, fragmented apophyseal bone, or fragmented annular tissue beyond the intervertebral disk space. The disk space is defined rostrally and caudally by the vertebral body endplates and peripherally by the outer edges of the vertebral ring apophyses, exclusive of osteophytic formations. The term *localized* contrasts with the term *generalized*, the latter being arbitrarily defined as greater than 50% (180°) of the periphery of the disk (28) (Fig 13).

Displacement, therefore, can occur only in association with disruption of the normal annulus or, as in the case of intravertebral herniation (Schmorl node) (Fig 14), a break in the vertebral body endplate. Since details of the integrity of the annulus are often unknown, the diagnosis of herniation is usually made by observation of displacement of disk material beyond the edges of the ring apophyses that is localized, meaning less than 50% (180°) of the circumference of the disk.

Localized displacement in the axial (horizontal) plane can be focal, signifying less than 25% of the disk circumference, or broad based, meaning between 25% and 50% of the disk circumference. Presence of disk tissue circumferentially (50%–100%) beyond the edges of the ring apophyses may be called bulging and is not considered a form of herniation.

A disk may have more than one herniation. The term *herniated disk* does not imply any knowledge of etiology, relation to symptoms, prognosis, or need for treatment. When data are sufficient to make the distinction, a herniated disk may be more specifically characterized as protruded or extruded. These distinctions are based on the shape of the displaced material. Protrusion is present if the greatest distance, in any plane, between the edges of the disk material beyond the disk space is less than the distance between the edges of the base in the same plane (Fig 15).

Figure 14



Figure 14: (a) Sagittal T1-weighted SE (450/15) and (b) sagittal midline T2-weighted fast SE (4000/120) images through lumbar spine. There is intrabody herniation into anterior superior aspect of L5 vertebral body and type II degenerative marrow change surrounding this herniation (arrow).

Figure 15

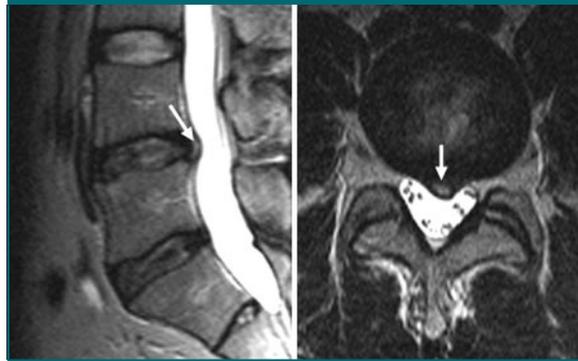


Figure 15: Protruded herniated disk. Sagittal (left) and transverse (right) T2-weighted fast SE (4000/120) images through lower lumbar spine and L4-5 disk level. Note convex posterior margin on left image (arrow). Protruded disk herniation is characterized by broader base than the extension of disk material beyond the disk space (arrow).

Extrusion is present when, in at least one plane, any one distance between the edges of the disk material beyond the disk space is greater than the distance between the edges of the base in the same plane or when no continuity exists between the disk material beyond the disk space and that within the disk space (Fig 16). Extrusion may be further specified as sequestration if the displaced disk material has lost completely any continuity with the parent disk. The term *migration* may be used to signify displacement of disk material away from

the site of extrusion, regardless of whether it is sequestered or not (Fig 17).

Herniated disks in the craniocaudal (vertical) direction through a break in the vertebral body endplate are referred to as intravertebral herniations (Fig 14). Nonacute Schmorl-node intrabody herniations are common spinal abnormalities regarded as incidental observations. They have been reported in 38%–75% of the population (69,70). While intrabody herniations may occur secondary to endplate weakness related to bone dysplasia, neoplasms, infec-

tions, or any process that weakens the endplate or the underlying bone, most intrabody herniations probably form after axial loading trauma, with preferential extrusion of nuclear material through the vertebral endplate rather than an intact and normal annulus fibrosis. It has been suggested that asymptomatic intrabody herniations may be traceable to a specific occurrence of acute nonradiating low back pain in the patient's history, which supports the concept that intrabody herniations (Schmorl nodes) occur through sites of endplate fracture. Type I vertebral body marrow changes have been described

surrounding the acute interbody herniations (71).

Stenosis

Spinal stenosis was defined in 1975 as any type of narrowing of the spinal canal, nerve root canals, or intervertebral foramina (72). Two broad groups have been defined: acquired (usually related to degenerative changes) and congenital or developmental. Developmental stenosis can be exacerbated by superimposed acquired degenerative changes. In the acquired type, there has been no association between the severity of pain and the degree of stenosis. The most

common symptoms are sensory disturbances in the legs, low back pain, neurogenic claudication, weakness, and relief of pain by bending forward. The imaging changes are in general more extensive than expected from the clinical findings (73). Patients with symptoms referable to spinal stenosis tend to have narrower spines than asymptomatic patients. While there does appear to be a correlation between cross-sectional area and midsagittal measurements in patients with symptomatic spinal stenosis, absolute values and correlation between measurements and symptoms appear to be lacking. The degree of stenosis is not static, and extension worsens the degree of central and foraminal stenosis by 11%, while flexion appears to improve it by an average of 11%. Segmental instability, which can cause static and dynamic stenosis, is considered a cause of low back pain but is poorly defined (74). Some evidence suggests that disk degeneration, narrowing of the spinal canal, and degenerative changes in the facets and spinal ligaments contribute to stenosis and that instability increases with age. Unfortunately, there do not appear to be reliable prognostic imaging findings that would correlate with surgical success or even whether patients would benefit from surgery (75).

Figure 16

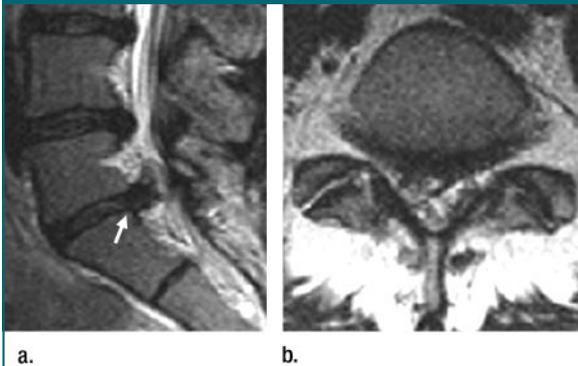


Figure 16: Extruded disk herniation. (a) Right parasagittal and (b) transverse T2-weighted fast SE images (4000/120) through lumbar spine and L5-S1 disk space. Extruded disk herniations are characterized by a base that is narrower than the distance of disk material extension beyond the disk space (arrow).

Figure 17

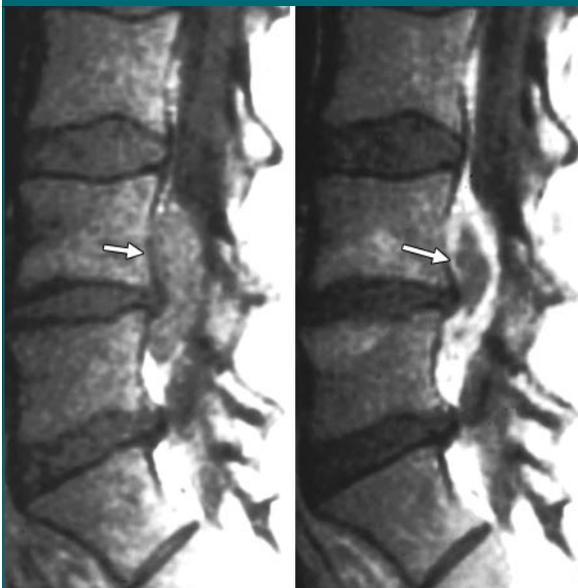


Figure 17: Extruded disk and free fragment. Sagittal T1-weighted SE (450/17) before (left) and after (right) contrast enhancement through lower lumbar spine. Note large extruded L4-5 disk fragment that has migrated superiorly behind the body of L4 (arrow). After administration of paramagnetic contrast media, there is peripheral enhancement of this mass, which represents granulation tissue within the epidural fibrosis surrounding nonenhancing central core of disk material.

Technical Considerations for MR Imaging

Traditional clinical imaging has emphasized orthogonal T1- and T2-weighted imaging for morphologic assessment of the discovertebral complex. These sequences also provide an evaluation of the signal intensity changes associated with degenerative disk disease. Fast SE T2-weighted images have replaced conventional T2-weighted images because of their shorter acquisition times, but they provide no increased diagnostic advantage. Short inversion time inversion-recovery or fat-suppressed T2-weighted images have been added by many groups, ours included, because it is believed they are more sensitive to marrow and soft-tissue changes.

While these standard sequences remain the mainstay of diagnostic imaging of the spine, new techniques continue to

be evaluated in hopes of providing stronger correlation between imaging findings and patient symptoms. The utility of many of these techniques for the routine evaluation of degenerative disk disease remains unknown, and the number of subjects in which they have been evaluated remains small. Nevertheless, these approaches may be important for redefining the direction of spinal imaging away from strictly anatomic one to one that combines more physiologic and functional information (76). Techniques that have been evaluated to greater or lesser degrees of success include assessment of spinal motion (dynamic imaging, kinetic assessment, or axial loading), diffusion imaging (water or contrast agents), MR neurography, spectroscopy, functional MR of the spinal cord, and ultrashort echo-time imaging. Of the variety of techniques available, only MR neurography and dynamic imaging have expanded beyond the experimental phase and have demonstrated specific clinical utility (albeit in niche or select populations).

Dynamic Imaging

The utility of dynamic spine MR remains unclear, in part due to the varied methods used. Methods to date include axial loading in the supine position by means of a harness that is attached to a non-magnetic compression footplate with nylon straps that can be tightened or use of an upright open MR system that allows flexion and extension imaging (77,78). Dynamic MR has been used to evaluate the occurrence of occult herniations, which may not be visible or be less visible when the patient is supine, to measure motion between spinal segments, and to measure the canal or foraminal diameter when subjected to axial loading (76,78–81). Hiwatashi et al (80) evaluated 200 patients with clinical symptoms of spinal stenosis and found 20 patients with detectable differences in caliber of the dural sac on routine and axial loaded studies. In five of these selected patients, all three neurosurgeons involved in the clinical evaluation changed their treatment decisions from conservative to decompressive surgery. While a small subset of patients may

benefit from this type of evaluation, the benefit appears small for the added machine time and patient discomfort.

Neurography

A large and varied literature exists concerning the use of MR neurography for the evaluation of peripheral nerves, including brachial and lumbar plexi. Thin-section MR neurography uses a high-resolution T1 imaging for anatomic detail and fat-suppressed T2-weighted or short inversion time inversion-recovery imaging to show abnormal nerve hyperintensity. Several reviews exist on this subject (82–84). The technique is capable of depicting a wide variety of pathologic conditions involving the sciatic nerve, such as compression, trauma, hypertrophy, neuroma, and tumor infiltration (85,86). MR neurography demonstrating piriformis syndrome (piriformis muscle asymmetry and sciatic nerve hyperintensity) has 93% specificity and 64% sensitivity (87).

Ultrashort Echo-time Imaging

Typical clinical MR imaging does not allow evaluation of tissues with very short relaxation times, since echo times are on the order of 8–15 msec. Ultrashort echo-time sequences have been preliminarily evaluated for a number of tissues, including the spine. These sequences have echo times as short as 0.08 msec. The images show normal contrast enhancement, with high signal intensity from longitudinal ligaments, endplate, and interspinous ligaments (88–90).

Diffusion

Several authors have evaluated the apparent diffusion coefficient (ADC) in normal and degenerated intervertebral disks. Antoniou et al (91) evaluated the ADC of cadaveric human disks related to matrix composition and matrix integrity by using a stimulated echo sequence. They found the ADCs in healthy subjects were significantly greater in the nucleus pulposus than in the annulus fibrosis. The ADCs were noted to generally decrease with degeneration grade and age in the nucleus. A similar correlation of ADC measurements and annu-

lar degeneration was not found. The most notable correlations were observed between the ADCs of nucleus pulposus and the water and glycoaminoglycan contents. Kealey et al (92) evaluated 39 patients with multishot SE echo-planar technique. They found a significant decrease in ADC of degenerated disks compared with that of normal disks. Kurunlahti et al (93) evaluated the ADC of disk and lumbar magnetic resonance angiograms in 37 asymptomatic volunteers. The lumbar artery status correlated with the diffusion values within the disks, suggesting that impaired blood flow may play an important role in disk degeneration. Kerttula et al (94) compared disk ADC values in normal controls with those in patients with prior compression fractures (at least 1 year previously) and found ADC values in x and y directions decreased in degenerated disks and in disks of normal signal intensity in the trauma area.

Diffusion-tensor imaging has been evaluated for imaging of the annulus fibrosus (95) and potentially for imaging defects or disruptions within the annulus (76). Differences in diffusion have been demonstrated for the intervertebral disk in compressed versus uncompressed states (96).

Intravenous contrast enhancement may also be used to assess diffusion into the intervertebral disk. Normal disks slowly enhance after contrast material injection, which may be as much as 36% in animal models. This enhancement is modified by the type of contrast agent (ionic vs nonionic) and molecular weight (97,98). Ionic material diffuses less rapidly into the disk than does nonionic media. Degenerated disks with decreased glycoaminoglycan have more intense and rapid enhancement (99). Disk enhancement has been documented in normal and degenerated human lumbar disks (100).

Symptoms

The etiology of symptoms in patients with degenerative disk disease is diverse, and there is often ambiguity in the diagnosis (101). The symptom com-

plexes are more often characterized by variability and change rather than predictability and stability (102).

The most common symptom is pain. Anatomic areas of the spine can serve as sites of pain generation through intrinsic innervation or acquired innervation as a product of soft-tissue repair. Mechanisms, which often act in combination, include (a) instability with associated disk degeneration, facet hypertrophy, or arthropathy; (b) mechanical compression of nerves by bone, ligament, or disk material; and (c) biochemical mediators of inflammation and/or pain.

It is important to reemphasize that disk degeneration per se is not painful, and in fact has a very high prevalence in the asymptomatic population. In addition, imaging findings of degenerative disk disease do not help predict a subsequent symptom development over time (103).

Mechanical compression or deformity of nerve roots as a cause of pain or nerve dysfunction is the classic concept related to displacement and effacement of neural tissue by disk herniation and dates to the observation of Mixter and Barr (104). Similar mechanical compression or traction mechanisms may be involved with instability or stenosis. A variety of morphologic changes occur in the nerve root with compression, including venous stasis, edema, and ultimately intraneural and perineural fibroses. Compression-induced impairment of both arterial and venous supply is one mechanism for nerve root dysfunction. Intraneural edema can occur even at low compression pressure levels (105). Mechanical compression itself may also be capable of producing changes in nerve impulses, which could be interpreted by the central nervous system as pain (106). However, the concept of neural compression by itself is inadequate to explain part or all of many symptom complexes.

The perplexing clinical scenario of patients who complain of incapacitating back pain, but may have no overt morphologic abnormality, has given rise to the concept of the disk as a pain generator. This was classically described by

Crock (107) as "chronic internal disc disruption syndrome" (108,109). Many different names have been given to this idea, which becomes more confusing when combined with the various diagnostic tests that are used in an attempt to diagnose this protean syndrome. Additional terms in the literature include internal annular tear, internal disk disruption, black disk disease, and discogenic pain. In a normal human lumbar disk, nerve endings can be found only in the periphery of the annulus, and the pain fibers are part of the sympathetic chain via the sinuvertebral nerve (110–112). This innervates the outer layer of the annulus fibrosis. However, in very degenerated disks, nerves may even penetrate into the nucleus pulposus (113). Potentially, stimulation of these fibers can occur not only from direct disruption and mechanical pressure on the annulus but also from various breakdown products of the nucleus pulposus or secondarily upregulated inflammatory mediators. Discography has been cited as the reference standard for the diagnosis of discogenic pain, but what it means in terms of patient care has never been prospectively tested.

The concept of disk tissue producing an inflammatory response is not new, but has become more sophisticated and targeted with the application of monoclonal antibody technology, and other assay techniques (114) demonstrated chemical radiculitis, which was thought related to nuclear material and its glycoproteins, as being highly irritant to nerve tissue. McCarron et al (115), using a dog model, demonstrated in 1987 that autogenous placement of nucleus pulposus into the epidural space caused acute and chronic inflammatory reaction, with influx of histocytes and fibroblasts. Kayama et al (116) and Olmarker et al (117) demonstrated that nucleus pulposus applied to spinal nerves induces a wide variety of functional, vascular, and morphologic abnormalities, often followed by intraradicular fibrosis and neural atrophy. Nucleus pulposus can cause an inflammatory reaction with leukotaxis and increased vascular permeability (118). Direct placement of nuclear material is

not necessary in animal models to induce an inflammatory response, but simply an incision of the annulus fibrosus can produce morphologic and functional changes in the adjacent nerves, such as increased capillaries and reduced nerve conduction velocities (116), with the presumed mechanism of disk material leakage into the epidural space. Monoclonal antibody staining of disk material has shown that the cells demonstrate an immunophenotype of inflammatory response, that is, macrophages (119). As a manifestation of this inflammatory response, higher systemic plasma levels of C-reactive protein have been found in patients with sciatica versus healthy controls (120).

Multiple studies have demonstrated vascularized granulation tissue surrounding the cartilage component of disk herniations (121,122), which correspond to the common enhanced MR findings of peripheral enhancement surrounding nonenhancing lumbar disk extrusions in unoperated patients. Blood vessels have been demonstrated in up to 91% of herniations, being most prevalent with disk sequestrations (123). Gronblad et al (124), using monoclonal antibodies, evaluated the types of inflammatory cells found with disk herniations and found them to be dominated by macrophages. There was also evidence of IL-1 β expression, an important proinflammatory cytokine.

Disk cells are also capable of expressing other proinflammatory substances, such as TNF- α , which can produce radicular morphologic abnormalities similar to those seen with nucleus pulposus application (125). TNF- α is overexpressed in degenerated disks and is a proinflammatory cytokine affecting matrix metalloproteinases (MMP) expression and increasing prostaglandin E₂. Weiler et al (126) demonstrated TNF- α in cross-sections of human disks and found synthesis of TNF- α in annular and disk regions, increased TNF- α with symptomatic disk disease, and TNF- α expression associated with increasing disk degeneration. Olmarker and Rydevik (127) showed that inhibition of TNF- α prevented thrombus formation and intraneural edema and reduced

nerve conduction velocity. This set the stage for an open label trial of anti-TNF therapy in patients with sciatica (128–130). Infliximab (Remicade; Centocor, Malvern, Pa), a chimeric monoclonal human and mouse antibody, inhibits TNF- α -induced infiltration of leukocytes to the site of injury. A single infusion of infliximab produced a rapid beneficial effect on pain, which persisted over 1 year, at the 3 mg/kg dose level. Sustained improvement was also demonstrated with the subcutaneous injection of another anti-TNF agent, etanercept (Enbrel; Immunex-Amgen, Thousand Oaks, Calif) (128). While intriguing, the off-label use of these TNF inhibitors is not recommended, since little data are currently available and only a very small number of patients have been treated. This trial does point out the direction of research for treatment of disk disease, with specific targeting of inflammatory pathways.

A wide variety of inflammatory agents are capable of expression from both migratory macrophages into the site of herniation and directly from stimulated chondrocytes (131). Burke et al (132) found increased levels of IL-6, IL-8, prostaglandin E2, and monocyte chemoattractant protein-1 in disk extracts of patients undergoing fusion for discogenic pain. Monocyte chemoattractant protein-1 is a CC chemokine that contributes to the activation and recruitment of macrophages and is expressed by chondrocytes that are stimulated by other cytokines and some MMPs. Disk tissue is biologically active and can respond to a proinflammatory stimulus by secreting IL-6, IL-8, and prostaglandin E2, but not TNF- α . In a rabbit disk herniation model, however, Yoshida et al (133) demonstrated infiltrating macrophages at day 3 postoperatively, with intervertebral disk cells producing TNF- α and IL-1 β on day 1 and monocyte chemoattractant protein-1 on day 3. In a mouse-derived coculture system of disk material and macrophages, Kato et al (134) also demonstrated upregulation of TNF- α messenger RNA and protein expression as the first point of the inflammatory cascade. The TNF- α -dependent glyco-

protein, TNF- α -stimulated gene-6 (TSG-6), which is found in inflammatory diseases of related connective tissues, has been demonstrated in 98% of disk herniations in one series (135).

Another component of the inflammatory response involves the matrix-degrading enzymes called MMPs. There are approximately 25 MMPs in five classes based on the specificity of their substrate. These enzymes degrade the extracellular matrix at physiologic pH levels. They are released by resident cells such as fibroblasts, macrophages in herniated disks, and chondrocytes from protrusions and nonherniated disk (136). MMPs can play a direct role in disk degeneration by causing matrix proteolysis and disk resorption and have an indirect role in angiogenesis. MMPs involved in disk degeneration include MMP-1 (collagenase); MMP-3 (stromelysin-1); MMP-9 (gelatinase B); and MMPs-2, -7, -8, and -13 (137). Cells within granulation tissue in disk herniations express MMP-1 and MMP-3 (138,139). MMP-3, but not MMP-7 (matrilysin), appears necessary for disk resorption, although the mechanism may be indirect and correlates with macrophage infiltration (140). The angiogenic properties are more indirect, with endothelial cell migration occurring only after a proteolytic reaction produced by MMP-3 (141). Cytokines leading to MMP production within the disk herniation may then result in angiogenesis and disk resorption. Given the wide variety of MMPs present, it is likely that there is a cascade of interacting proteases for different components of the disk matrix involved in disk resorption and degeneration.

Many other molecules have also shown to be present in degenerated or herniated disks that may play additional roles in the inflammatory cascade, such as intercellular adhesion molecule-1, fibroblast growth factor, and vascular endothelial growth factor (142,143). These two latter agents contribute to neoangiogenesis. Vascular endothelial growth factor appears to require TNF- α for induction. Additionally, it is a potent inducer of plasmin and results in activation of a variety of MMPs. Interaction

between the vascular endothelial growth factor and MMPs could promote disk matrix degeneration, as well as neovascularization of herniations.

Nerve fibers have been identified in the outer third of the annulus in the normal state, but may extend into the inner annulus and nucleus pulposus, accompanied by blood vessels, in chronic back pain patients (113). These nerves also stain for substance P, a putative nociceptive neurotransmitter, along with calcitonin gene-related peptide and vasoactive intestinal peptide. These small nonmyelinated nerve fibers grow into the disk in areas with local production of nerve growth factor, which is produced by the neoangiogenesis of the disk material (113). Coupled with nerve ingrowth and angiogenesis is the production of inflammation by liberation of the potent inflammatory agent phospholipase A2, which catalyzes the hydrolysis of phosphoglyceride, an important membrane constituent (144,145). This production of phospholipase A2 is induced by, among other signals, the presence of IL-1 and TNF- α , with the subsequent upregulation of the arachadonic acid cascade, which produces prostaglandin E2 and leukotrienes (146,147). Prostaglandin E2 production has the critical rate-limiting enzyme cyclooxygenase-2, which appears to be primarily upregulated during inflammation (148). The other branch of the arachadonic acid cascade is the production of leukotrienes by means of the enzyme lipoxygenase. A bell-shaped pain behavior dose response curve has been demonstrated for intraneural injection of TNF- α and IL-1 β in a rat model, peaking at doses equivalent to those of endogenous cytokines released locally after nerve injury. An increase in perineural macrophages was also observed, particularly for IL-1 β (149).

Sensory and motor deficits would appear to be the result of both a combination of mechanical deformation and the presence of inflammation. A variety of mechanisms and mediators are involved in the inflammatory side of the equation, and a brief general overview is attempted in Figure 18. Clearly, the etiology of pain in degenerative disease

is much more complex than a simple mechanical explanation, and work on these other factors will hopefully bring us a greater understanding of the relationship between morphologic alteration and clinical symptoms.

Importance of Imaging Findings

The role of an imaging test is to provide accurate morphologic information and influence therapeutic decision making (150). A necessary component, which connects these two purposes, is accurate natural history data.

Modern imaging has made important strides in supporting the first goal, accurate morphologic information. Not only are morphologic changes depicted in ever-increasing anatomic detail, but additional information from the imaging study has been made available that is helping us to understand cellular and biochemical alterations. The ability to better characterize these alterations should provide a means of more accu-

rately stratifying patient changes that may allow a more accurate understanding of etiology.

Any study looking at the natural history of degenerative disk disease, prognostic value of imaging, or its effect on therapeutic decision making will be confounded by the high prevalence of morphologic change in the asymptomatic population (151–153). A 20%–28% of asymptomatic patients demonstrate disk herniations, and the majority have evidence of additional degenerative disk disease (151–153). These findings are not only nonpredictive at the moment, but prospectively as well. In a 7-year follow-up of the Borenstein et al (103) original patient group, the original MR findings were not predictive of the development or duration of low back pain.

As to natural history, some information is available. Degenerative disk space narrowing, facet disease, and stenosis tend to slowly progress over time. Eventual stabilization of the three-joint discovertebral complex is thought to be part of the natural history of degenerative disease, and it is assumed to be accompanied by a decrease in pain. These impressions, however, are anecdotal and have not been tested by a formal natural history study. Some findings, such as disk herniation and degenerative marrow changes, are known to change. Multiple studies in which computed tomography or MR imaging has been used have shown that the size of disk herniations, especially larger ones, can reduce dramatically in patients undergoing conservative treatment (154,155).

In a study of symptomatic patients, the prevalence of disk herniation in patients with low back pain and those with radiculopathy at presentation was similar (156). There was a higher prevalence of herniation, 57% in patients with low back pain and 65% in patients with radiculopathy, than the 20%–28% prevalence reported in asymptomatic series (152,153). Disks characterized as extruded showed more marked regression in patients with both low back pain and radiculopathy. In general, one-third of patients with disk herniation at presentation had significant resolution or

disappearance by 6 weeks and two-thirds by 6 months (155,156). The type, size, and location of herniation at presentation and changes in herniation size and type over time did not correlate with outcome. In fact, the presence of a herniation at a MR was a positive prognostic finding (156).

Interestingly, not only do disk herniations have a tendency to regress, but also new or larger ones may appear after the onset of symptoms. In this study, 13% of patients in this symptomatic series developed new or larger disk herniations over a 6-week period. In looking at patients with low back pain or radiculopathy, MR did not have additive value over clinical assessment. No prognostic sign that might alter treatment versus clinical assessment alone was identified. The size and type of disk herniation and location and presence of nerve root compression, significant in terms of morphologic alteration, were not related to patient outcome. Likewise, the presence or absence of stenosis, facet disease, or degenerative marrow changes did not correlate with patient outcome (156).

This lack of prognostic value also appears to apply to the conservative management of spinal stenosis. There do not appear to be reliable prognostic imaging findings that would correlate with surgical success or even whether patients would benefit from surgery and spinal stenosis (75,157). A study of the qualitative morphologic features of the spinal canal dimensions and herniated disks has not proved helpful in predicting outcomes in patients with back pain and sciatica. Demographic and clinical features appear to predict outcome of nonsurgical treatment, whereas morphometric features of disk herniation and spinal canal are more powerful predictors of surgical outcome (158).

Degenerative marrow changes may also change over time. In all three types, there is always evidence of associated degenerative disk disease at the level of involvement. Type I changes may revert to normal or convert to type II changes, with time suggesting some stabilization of the degenerative process. Type II changes tend to be more stable but may

Figure 18

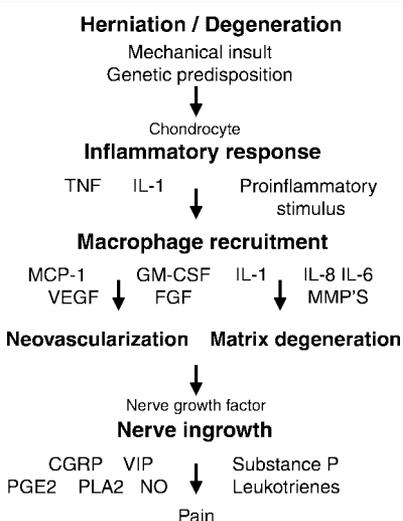


Figure 18: Flowchart of hypothetical inflammatory cascade for degenerative disk disease. *CGRP* = calcitonin gene-related peptide, *FGF* = fibroblast growth factor, *GM-CSF* = granulocyte-macrophage colony-stimulating factor, *MCP* = monocyte chemoattractant protein-1, *NO* = nitric oxide, *PGE2* = prostaglandin E2, *PLA2* = phospholipase A2, *VEGF* = vascular endothelial growth factor, *VIP* = vasoactive intestinal peptide.

convert to type I or a mixed combination of types I and II. When changes do occur in type II marrow, they are usually associated with evidence of additional or accelerated degeneration or a superimposed process such as infection or trauma.

The clinical importance of marrow changes associated with degenerative disk disease remains unclear. Type I changes seem to be associated with a higher prevalence of active low back pain symptoms. The exact etiologic mechanism or mechanisms, while unknown, have been thought related to some type of unusual stresses, micro- or macroinstability or microtrauma. Some studies of discography in patients with degenerative marrow changes have suggested that type I marrow changes are invariably associated with painful disks (159–162). Surgical studies have suggested that patients with type I marrow changes who undergo fusion for low back pain do better than those without endplate changes or type II patterns (159). The hypothesis is that type I degenerative marrow changes are related to or are indicators of some degree of instability. Authors of a surgical study looking at the prognostic value of type I marrow changes related to surgical outcome demonstrated that persistence of type I marrow changes after fusion was associated with significantly worse outcome (163). The authors speculate that type I changes may not only be an important criterion for surgery, but type I change disappearance may be an indicator of successful fusion and stabilization. Additional evidence to support that these changes are a reflection of a more active process related to microtrauma and instability is a surgical study that found that in the overwhelming majority of patients with type I marrow changes who undergo fixation and fusion, the marrow changes will convert to a normal marrow signal intensity or type II changes, with good clinical results (159).

Surgery

Degeneration of the intervertebral disk complex is a process that begins early in

life and is a consequence of a variety of genetic, physiologic, and environmental factors, as well as normal aging. Given the ubiquitous nature of the process and its high prevalence in both symptomatic and asymptomatic individuals, the jump from identifying an anatomic derangement to proposing a symptom complex must be made with caution (2). There is an opportunity for imaging to further our understanding of the process.

What separates individuals with dramatic morphologic findings who have no symptoms from individuals with identical alterations who do? Understanding the relationship of etiologic factors, the morphologic alterations, which can be characterized at imaging, and the mechanisms of pain production and their interactions in the production of symptoms will require more accurate and reproducible stratification of patient cohorts. This may be a strong suit of imaging, the phenotyping of morphologic alterations to compare with the emerging genotyping work relative to etiology and clinical manifestations. The ultimate translational goal is the integration of this newfound understanding into the therapeutic decision-making process.

References

- Pritzker KP. Aging and degeneration in the lumbar intervertebral disc. *Orthop Clin North Am* 1977;8:66–77.
- White AA 3rd, Gordon SL. Synopsis: workshop on idiopathic low-back pain. *Spine* 1982;7:141–149.
- Adams MA, Hutton WC. Gradual disc prolapse. *Spine* 1985;10:524–531.
- Adams MA, Hutton WC. Prolapsed intervertebral disc: a hyperflexion injury 1981 Volvo Award in Basic Science. *Spine* 1982;7:184–191.
- Gordon SJ, Yang KH, Mayer PJ, Mace AH Jr, Kish VL, Radin EL. Mechanism of disc rupture: a preliminary report. *Spine* 1991;16:450–456.
- McNally DS, Adams MA, Goodship AE. Can intervertebral disc prolapse be predicted by disc mechanics? *Spine* 1993;18:1525–1530.
- Shirazi-Adl A. Strain in fibers of a lumbar disc: analysis of the role of lifting in producing disc prolapse. *Spine* 1989;14:96–103.
- Adams MA, Freeman BJ, Morrison HP, Nelson IW, Dolan P. Mechanical initiation of intervertebral disc degeneration. *Spine* 2000;25:1625–1636.
- Hurri H, Karppinen J. Discogenic pain. *Pain* 2004;112:225–228.
- Kauppila LI, McAlindon T, Evans S, Wilson PW, Kiel D, Felson DT. Disc degeneration/back pain and calcification of the abdominal aorta: a 25-year follow-up study in Framingham. *Spine* 1997;22:1642–1647.
- Kresina TF, Malesud CJ, Moskowitz RW. Analysis of osteoarthritic cartilage using monoclonal antibodies reactive with rabbit proteoglycan. *Arthritis Rheum* 1986;29:863–871.
- Battie MC, Videman T, Gibbons LE, Fisher LD, Manninen H, Gill K. 1995 Volvo Award in clinical sciences: determinants of lumbar disc degeneration—a study relating lifetime exposures and magnetic resonance imaging findings in identical twins. *Spine* 1995;20:2601–2612.
- Hestbaek L, Iachine IA, Leboeuf-Yde C, Kyvik KO, Manniche C. Heredity of low back pain in a young population: a classical twin study. *Twin Res* 2004;7:16–26.
- Annunen S, Paasilta P, Lohiniva J, et al. An allele of COL9A2 associated with intervertebral disc disease. *Science* 1999;285:409–412.
- Marini JC. Genetic risk factors for lumbar disk disease. *JAMA* 2001;285:1886–1888.
- Paasilta P, Lohiniva J, Goring HH, et al. Identification of a novel common genetic risk factor for lumbar disk disease. *JAMA* 2001;285:1843–1849.
- Watanabe H, Nakata K, Kimata K, Nakanishi I, Yamada Y. Dwarfism and age-associated spinal degeneration of heterozygote cmd mice defective in aggrecan. *Proc Natl Acad Sci U S A* 1997;94:6943–6947.
- Nelson CL, Janecki CJ, Gildenberg PL, Sava G. Disk protrusions in the young. *Clin Orthop Relat Res* 1972;88:142–150.
- Varlotta GP, Brown MD, Kelsey JL, Golden AL. Familial predisposition for herniation of a lumbar disc in patients who are less than 21 years old. *J Bone Joint Surg Am* 1991;73:124–128.
- Matsui H, Terahata N, Tsuji H, Hirano N, Naruse Y. Familial predisposition and clustering for juvenile lumbar disc herniation. *Spine* 1992;17:1323–1328.
- Scapinelli R. Lumbar disc herniation in eight siblings with a positive family history

- for disc disease. *Acta Orthop Belg* 1993;59:371-376.
22. Solovieva S, Leino-Arjas P, Saarela J, Luoma K, Raininko R, Riihimaki H. Possible association of interleukin 1 gene locus polymorphisms with low back pain. *Pain* 2004;109:8-19.
 23. Quinet RJ, Hadler NM. Diagnosis and treatment of backache. *Semin Arthritis Rheum* 1979;8:261-287.
 24. Bonneville JF, Dietemann JL. Imaging in sciatica [in French]. *Rev Prat* 1992;42:554-566.
 25. Brant-Zawadzki MN, Jensen MC, Obuchowski N, Ross JS, Modic MT. Interobserver and intraobserver variability in interpretation of lumbar disc abnormalities: a comparison of two nomenclatures. *Spine* 1995;20:1257-1263.
 26. Breton G. Is that a bulging disk, a small herniation or a moderate protrusion? *Can Assoc Radiol J* 1991;42:318.
 27. Fardon DF, Balderston RA, Garfin SR, Nasca RJ, Pinkerton S, Salib RM. Disorders of the spine: a coding system for diagnoses. Philadelphia, Pa: Hanley & Belfus, 1991; 20-22.
 28. Milette PC. Reporting lumbar disk abnormalities: at last, consensus! *AJNR Am J Neuroradiol* 2001;22:428-429.
 29. Czervionke LF. Lumbar intervertebral disc disease. *Neuroimaging Clin N Am* 1993;3:465-485.
 30. Modic MT, Herfkens RJ. Intervertebral disk: normal age-related changes in MR signal intensity. *Radiology* 1990;177:332-333.
 31. Sether LA, Yu S, Houghton VM, Fischer ME. Intervertebral disk: normal age-related changes in MR signal intensity. *Radiology* 1990;177:385-388.
 32. Resnick D, Niwayama G. Degenerative disease of the spine. Philadelphia, Pa: Saunders, 1995; 1372-1462.
 33. Schmorl G, Junghanns H. The human spine in health and disease. 2nd American ed. New York, NY: Grune & Stratton, 1971; 141-148.
 34. Nathan H. Osteophytes of the vertebral column: an anatomical study of their development according to age, race, and sex with consideration as to their etiology and significance. *J Bone Joint Surg* 1962;44:243-268.
 35. Sauser DD, Goldman AB, Kaye JJ. Discogenic vertebral sclerosis. *J Can Assoc Radiol* 1978;29:44-50.
 36. Twomey L, Taylor J. Age changes in lumbar intervertebral disc. *Acta Orthop Scand* 1985;56:496-499.
 37. Twomey LT, Taylor JR. Age changes in lumbar vertebrae and intervertebral discs. *Clin Orthop Relat Res* 1987;224:97-104.
 38. Thalgott JS, Albert TJ, Vaccaro AR, et al. A new classification system for degenerative disc disease of the lumbar spine based on magnetic resonance imaging, provocative discography, plain radiographs and anatomic considerations. *Spine J* 2004;4(6 suppl):167S-172S.
 39. Iida T, Abumi K, Kotani Y, Kaneda K. Effects of aging and spinal degeneration on mechanical properties of lumbar supraspinous and interspinous ligaments. *Spine J* 2002;2:95-100.
 40. Kirkaldy-Willis WH, Wedge JH, Yong-Hing K, Reilly J. Pathology and pathogenesis of lumbar spondylosis and stenosis. *Spine* 1978;3:319-328.
 41. Adams P, Eyre DR, Muir H. Biochemical aspects of development and ageing of human lumbar intervertebral discs. *Rheumatol Rehabil* 1977;16:22-29.
 42. Brown MD. The pathophysiology of disc disease. *Orthop Clin North Am* 1971;2:359-370.
 43. Lipson SJ, Muir H. Experimental intervertebral disc degeneration: morphologic and proteoglycan changes over time. *Arthritis Rheum* 1981;24:12-21.
 44. Modic MT, Pavlicek W, Weinstein MA, et al. Magnetic resonance imaging of intervertebral disk disease: clinical and pulse sequence considerations. *Radiology* 1984;152:103-111.
 45. Majors AW, McDevitt CA, Silgalis I, Modic MT. A correlative analysis of T2, ADC and MT radio with water, hydroxyproline and GAG content in excised human intervertebral disk. New Orleans, La; Orthopedic Research Society, 1994; 116-120.
 46. Yu S, Houghton VM, Sether LA, Ho KC, Wagner M. Criteria for classifying normal and degenerated lumbar intervertebral disks. *Radiology* 1989;170:523-526.
 47. Friberg S, Hirsch C. Anatomical and clinical studies on lumbar disc degeneration. *Acta Orthop Scand* 1949;19:222-242.
 48. Hirsch C. Studies on the pathology of low back pain. *J Bone Joint Surg Br* 1959;41-B:237-243.
 49. Coventry MB. Anatomy of the intervertebral disk. *Clin Orthop Relat Res* 1969;67:9-15.
 50. Pfirrmann CW, Metzendorf A, Zanetti M, Hodler J, Boos N. Magnetic resonance classification of lumbar intervertebral disc degeneration. *Spine* 2001;26:1873-1878.
 51. Knutsson F. The vacuum phenomenon in the intervertebral discs. *Acta Radiol* 1942;23:173-179.
 52. Grenier N, Grossman RI, Schiebler ML, Yeager BA, Goldberg HI, Kressel HY. Degenerative lumbar disk disease: pitfalls and usefulness of MR imaging in detection of vacuum phenomenon. *Radiology* 1987;164:861-865.
 53. Bielecki DK, Sartoris D, Resnick D, Van Lom K, Fierer J, Haghighi P. Intraosseous and intradiscal gas in association with spinal infection: report of three cases. *AJR Am J Roentgenol* 1986;147:83-86.
 54. Bangert BA, Modic MT, Ross JS, et al. Hyperintense disks on T1-weighted MR images: correlation with calcification. *Radiology* 1995;195:437-443.
 55. Boyko OB, Burger PC, Shelburne JD, Ingram P. Non-heme mechanisms for T1 shortening: pathologic CT, and MR elucidation. *AJNR Am J Neuroradiol* 1992;13:1439-1445.
 56. Henkelman RM, Watts JF, Kucharczyk W. High signal intensity in MR images of calcified brain tissue. *Radiology* 1991;179:199-206.
 57. de Roos A, Kressel H, Spritzer C, Dalinka M. MR imaging of marrow changes adjacent to end plates in degenerative lumbar disk disease. *AJR Am J Roentgenol* 1987;149:531-534.
 58. Masaryk TJ, Boumpfrey F, Modic MT, Tamborrello C, Ross JS, Brown MD. Effects of chemonucleolysis demonstrated by MR imaging. *J Comput Assist Tomogr* 1986;10:917-923.
 59. Modic MT, Steinberg PM, Ross JS, Masaryk TJ, Carter JR. Degenerative disk disease: assessment of changes in vertebral body marrow with MR imaging. *Radiology* 1988;166:193-199.
 60. Ross JS, Robertson JT, Frederickson RC, et al. Association between peridural scar and recurrent radicular pain after lumbar discectomy: magnetic resonance evaluation. *ADCON-L European Study Group. Neurosurgery* 1996;38:855-861.
 61. Ross JS, Obuchowski N, Zepp R. The post-operative lumbar spine: evaluation of epidural scar over a 1-year period. *AJNR Am J Neuroradiol* 1998;19:183-186.
 62. Morrison JL, Kaplan PA, Dussault RG, Anderson MW. Pedicle marrow signal intensity changes in the lumbar spine: a manifestation of facet degenerative joint disease. *Skeletal Radiol* 2000;29:703-707.

63. Ulmer JL, Elster AD, Mathews VP, Allen AM. Lumbar spondylolysis: reactive marrow changes seen in adjacent pedicles on MR images. *AJR Am J Roentgenol* 1995; 164:429–433.
64. Harris RI, MacNab I. Structural changes in the lumbar intervertebral discs: their relationship to low back pain and sciatica. *J Bone Joint Surg Br* 1954;36-B:304–322.
65. Schellinger D, Wener L, Ragsdale BD, Patronas NJ. Facet joint disorders and their role in the production of back pain and sciatica. *RadioGraphics* 1987;7:923–944.
66. Doyle AJ, Merrilees M. Synovial cysts of the lumbar facet joints in a symptomatic population: prevalence on magnetic resonance imaging. *Spine* 2004;29(8):874–878.
67. Bastrup CI. On the spinous processes of the lumbar vertebrae and the soft tissue between them and on pathological changes in that region. *Acta Radiol Scan* 1933;14: 52–54.
68. Jacobson HG, Tausend ME, Shapiro JH, Poppel MH. The swayback syndrome. *Am J Roentgenol Radium Ther Nucl Med* 1958; 79:677–683.
69. Hilton RC, Ball J, Benn RT. Vertebral endplate lesions (Schmorl's nodes) in the dorsolumbar spine. *Ann Rheum Dis* 1976;35: 127–132.
70. Resnick D, Niwayama G. Intravertebral disk herniations: cartilaginous (Schmorl's) nodes. *Radiology* 1978;126:57–65.
71. Wagner AL, Murtagh FR, Arrington JA, Stallworth D. Relationship of Schmorl's nodes to vertebral body endplate fractures and acute endplate disk extrusions. *AJNR Am J Neuroradiol* 2000;21:276–281.
72. Arnoldi CC, Brodsky AE, Cauchoix J, et al. Lumbar spinal stenosis and nerve root entrapment syndromes: definition and classification. *Clin Orthop Relat Res* 1976; 115:4–5.
73. Amundsen T, Weber H, Lilleas F, Nordal HJ, Abdelnoor M, Magnaes B. Lumbar spinal stenosis: clinical and radiologic features. *Spine* 1995;20:1178–1186.
74. Inufusa A, An HS, Lim TH, Hasegawa T, Haughton VM, Nowicki BH. Anatomic changes of the spinal canal and intervertebral foramen associated with flexion-extension movement. *Spine* 1996;21:2412–2420.
75. ECRI. Treatment of degenerative lumbar spinal stenosis. I. Evidence report. Agency for Healthcare Research and Quality publication no. 01-E048 #32. Plymouth Meeting, Pa: ECRI, 2001.
76. Haughton V. Medical imaging of intervertebral disc degeneration: current status of imaging. *Spine* 2004;29:2751–2756.
77. Jinkins JR, Dworkin JS, Damadian RV. Upright, weight-bearing, dynamic-kinetic MRI of the spine: initial results. *Eur Radiol* 2005; 15(9):1815–1825.
78. Danielson B, Willen J. Axially loaded magnetic resonance image of the lumbar spine in asymptomatic individuals. *Spine* 2001; 26:2601–2606.
79. Kimura S, Steinbach GC, Watenpau DE, Hargens AR. Lumbar spine disc height and curvature responses to an axial load generated by a compression device compatible with magnetic resonance imaging. *Spine* 2001;26:2596–2600.
80. Hiwatashi A, Danielson B, Moritani T, et al. Axial loading during MR imaging can influence treatment decision for symptomatic spinal stenosis. *AJNR Am J Neuroradiol* 2004;25:170–174.
81. Tanaka N, An HS, Lim TH, Fujiwara A, Jeon CH, Haughton VM. The relationship between disc degeneration and flexibility of the lumbar spine. *Spine J* 2001;1:47–56.
82. Aagaard BD, Maravilla KR, Kliot M. Magnetic resonance neurography: magnetic resonance imaging of peripheral nerves. *Neuroimaging Clin N Am* 2001;11:viii, 131–146.
83. Filler AG, Maravilla KR, Tsuruda JS. MR neurography and muscle MR imaging for image diagnosis of disorders affecting the peripheral nerves and musculature. *Neurol Clin* 2004;22:643–682, vi–vii.
84. Maravilla KR, Bowen BC. Imaging of the peripheral nervous system: evaluation of peripheral neuropathy and plexopathy. *AJNR Am J Neuroradiol* 1998;19:1011–1023.
85. Moore KR, Tsuruda JS, Dailey AT. The value of MR neurography for evaluating extraspinal neuropathic leg pain: a pictorial essay. *AJNR Am J Neuroradiol* 2001;22: 786–794.
86. Ellegala DB, Monteith SJ, Haynor D, Bird TD, Goodkin R, Kliot M. Characterization of genetically defined types of Charcot-Marie-Tooth neuropathies by using magnetic resonance neurography. *J Neurosurg* 2005;102:242–245.
87. Filler AG, Haynes J, Jordan SE, et al. Sciatica of nondisc origin and piriformis syndrome: diagnosis by magnetic resonance neurography and interventional magnetic resonance imaging with outcome study of resulting treatment. *J Neurosurg* 2005;2:99–115.
88. Gatehouse PD, He T, Hughes SP, Bydder GM. MR imaging of degenerative disc disease in the lumbar spine with ultrashort TE pulse sequences. *MAGMA* 2004;16:160–166.
89. Hall-Craggs MA, Porter J, Gatehouse PD, Bydder GM. Ultrashort echo time (UTE) MRI of the spine in thalassaemia. *Br J Radiol* 2004;77:104–110.
90. Robson MD, Gatehouse PD, So PW, Bell JD, Bydder GM. Contrast enhancement of short T2 tissues using ultrashort TE (UTE) pulse sequences. *Clin Radiol* 2004;59:720–726.
91. Antoniou J, Demers CN, Beaudoin G, et al. Apparent diffusion coefficient of intervertebral discs related to matrix composition and integrity. *Magn Reson Imaging* 2004; 22:963–972.
92. Kealey SM, Aho T, Delong D, Barboriak DP, Provenzale JM, Eastwood JD. Assessment of apparent diffusion coefficient in normal and degenerated intervertebral lumbar disks: initial experience. *Radiology* 2005;235:569–574.
93. Kurunlahti M, Kerttula L, Jauhiainen J, Karppinen J, Tervonen O. Correlation of diffusion in lumbar intervertebral disks with occlusion of lumbar arteries: a study in adult volunteers. *Radiology* 2001;221:779–786.
94. Kerttula L, Kurunlahti M, Jauhiainen J, Koivula A, Oikarinen J, Tervonen O. Apparent diffusion coefficients and T2 relaxation time measurements to evaluate disc degeneration: a quantitative MR study of young patients with previous vertebral fracture. *Acta Radiol* 2001;42:585–591.
95. Hsu EW, Setton LA. Diffusion tensor microscopy of the intervertebral disc annulus fibrosus. *Magn Reson Med* 1999;41:992–999.
96. Chiu EJ, Newitt DC, Segal MR, Hu SS, Lotz JC, Majumdar S. Magnetic resonance imaging measurement of relaxation and water diffusion in the human lumbar intervertebral disc under compression in vitro. *Spine* 2001;26:E437–E444.
97. Perlewitz TJ, Haughton VM, Riley LHI, Nguyen-minh C, George V. Effect of molecular weight on the diffusion of contrast media into cartilage. *Spine* 1997;22:2707–2710.
98. Akansel G, Haughton VM, Papke RA, Censky S. Diffusion into human intervertebral disks studied with MR and gadoteridol. *AJNR Am J Neuroradiol* 1997;18:443–445.
99. Ibrahim MA, Haughton VM, Hyde JS. Effect of disk maturation on diffusion of low-molecular-weight gadolinium complexes:

- an experimental study in rabbits. *AJNR Am J Neuroradiol* 1995;16:1307-1311.
100. Nguyen-minh C, Houghton VM, Papke RA, An H, Censky SC. Measuring diffusion of solutes into intervertebral disks with MR imaging and paramagnetic contrast medium. *AJNR Am J Neuroradiol* 1998;19:1781-1784.
 101. Deyo RA. Practice variations, treatment fads, rising disability: do we need a new clinical research paradigm? *Spine* 1993;18:2153-2162.
 102. Von Korff M, Saunders K. The course of back pain in primary care. *Spine* 1996;21:2833-2837.
 103. Borenstein DG, O'Mara JW Jr, Boden SD, et al. The value of magnetic resonance imaging of the lumbar spine to predict low-back pain in asymptomatic subjects: a 7-year follow-up study. *J Bone Joint Surg Am* 2001;83-A:1306-1311.
 104. Mixter W, Barr JS. Rupture of the intervertebral disc with involvement of the spinal canal. *N Engl J Med* 1934;211.
 105. Olmarker K, Rydevik B, Holm S. Edema formation in spinal nerve roots induced by experimental, graded compression: an experimental study on the pig cauda equina with special reference to differences in effects between rapid and slow onset of compression. *Spine* 1989;14:569-573.
 106. Howe JF, Loeser JD, Calvin WH. Mechanosensitivity of dorsal root ganglia and chronically injured axons: a physiological basis for the radicular pain of nerve root compression. *Pain* 1977;3:25-41.
 107. Crock HV. Internal disc disruption: a challenge to disc prolapse fifty years on. *Spine* 1986;11:650-653.
 108. Thompson KJ. *Diagnostic lumbar disc injection*. 3rd ed. Philadelphia, Pa: Lippincott Williams & Wilkins, 2004; 87-90.
 109. Yonemura KS, Yuan H. The black disc. In: Benzel E, ed. *Spine surgery: techniques, complication avoidance and management*. 2nd ed. Philadelphia, Pa: Elsevier, 2004; 626-631.
 110. Edgar MA, Ghadially JA. Innervation of the lumbar spine. *Clin Orthop Relat Res* 1976; 115:35-41.
 111. Ashton IK, Roberts S, Jaffray DC, Polak JM, Eisenstein SM. Neuropeptides in the human intervertebral disc. *J Orthop Res* 1994;12:186-192.
 112. Fagan A, Moore R, Vernon Roberts B, Blumberg P, Fraser R. ISSLS prize winner: the innervation of the intervertebral disc—a quantitative analysis. *Spine* 2003; 28:2570-2576.
 113. Freemont AJ, Peacock TE, Goupille P, Hoyland JA, O'Brien J, Jayson MI. Nerve ingrowth into diseased intervertebral disc in chronic back pain. *Lancet* 1997;350:178-181.
 114. Marshall LL, Trethewie ER, Curtain CC. Chemical radiculitis: a clinical, physiological and immunological study. *Clin Orthop Relat Res* 1977;129:61-67.
 115. McCarron RF, Wimpee MW, Hudkins PG, Laros GS. The inflammatory effect of nucleus pulposus: a possible element in the pathogenesis of low-back pain. *Spine* 1987; 12:760-764.
 116. Kayama S, Konno S, Olmarker K, Yabuki S, Kikuchi S. Incision of the annulus fibrosus induces nerve root morphologic, vascular, and functional changes: an experimental study. *Spine* 1996;21:2539-2543.
 117. Olmarker K, Nordborg C, Larsson K, Rydevik B. Ultrastructural changes in spinal nerve roots induced by autologous nucleus pulposus. *Spine* 1996;21:411-414.
 118. Olmarker K, Blomquist J, Stromberg J, Nannmark U, Thomsen P, Rydevik B. Inflammatory properties of nucleus pulposus. *Spine* 1995;20:665-669.
 119. Kawaguchi S, Yamashita T, Yokogushi K, Murakami T, Ohwada O, Sato N. Immunophenotypic analysis of the inflammatory infiltrates in herniated intervertebral discs. *Spine* 2001;26:1209-1214.
 120. Le Gars L, Borderie D, Kaplan G, Berenbaum F. Systemic inflammatory response with plasma C-reactive protein elevation in disc-related lumbosciatic syndrome. *Joint Bone Spine* 2000;67(5):452-455.
 121. Doita M, Kanatani T, Harada T, Mizuno K. Immunohistologic study of the ruptured intervertebral disc of the lumbar spine. *Spine* 1996;21:235-241.
 122. Ito T, Yamada M, Ikuta F, et al. Histologic evidence of absorption of sequestration-type herniated disc. *Spine* 1996;21:230-234.
 123. Virri J, Gronblad M, Savikko J, et al. Prevalence, morphology, and topography of blood vessels in herniated disc tissue: a comparative immunocytochemical study. *Spine* 1996;21:1856-1863.
 124. Gronblad M, Virri J, Tolonen J, et al. A controlled immunohistochemical study of inflammatory cells in disc herniation tissue. *Spine* 1994;19:2744-2751.
 125. Igarashi T, Kikuchi S, Shubayev V, Myers RR. 2000 Volvo Award winner in basic science studies: exogenous tumor necrosis factor-alpha mimics nucleus pulposus-induced neuropathology—molecular, histologic, and behavioral comparisons in rats. *Spine* 2000;25:2975-2980.
 126. Weiler C, Nerlich AG, Bachmeier BE, Boos N. Expression and distribution of tumor necrosis factor alpha in human lumbar intervertebral discs: a study in surgical specimen and autopsy controls. *Spine* 2005;30:44-53.
 127. Olmarker K, Rydevik B. Selective inhibition of tumor necrosis factor-alpha prevents nucleus pulposus-induced thrombus formation, intraneural edema, and reduction of nerve conduction velocity: possible implications for future pharmacologic treatment strategies of sciatica. *Spine* 2001;26:863-869.
 128. Genevay S, Stingelin S, Gabay C. Efficacy of etanercept in the treatment of acute, severe sciatica: a pilot study. *Ann Rheum Dis* 2004;63:1120-1123.
 129. Karppinen J, Korhonen T, Malmivaara A, et al. Tumor necrosis factor-alpha monoclonal antibody, infliximab, used to manage severe sciatica. *Spine* 2003;28:750-753.
 130. Korhonen T, Karppinen J, Malmivaara A, et al. Efficacy of infliximab for disc herniation-induced sciatica: 1-year follow-up. *Spine* 2004;29:2115-2119.
 131. Takahashi H, Suguro T, Okazima Y, Motegi M, Okada Y, Kakiuchi T. Inflammatory cytokines in the herniated disc of the lumbar spine. *Spine* 1996;21:218-224.
 132. Burke JG, Watson RW, McCormack D, Dowling FE, Walsh MG, Fitzpatrick JM. Spontaneous production of monocyte chemoattractant protein-1 and interleukin-8 by the human lumbar intervertebral disc. *Spine* 2002;27:1402-1407.
 133. Yoshida M, Nakamura T, Sei A, Kikuchi T, Takagi K, Matsukawa A. Intervertebral disc cells produce tumor necrosis factor alpha, interleukin-1beta, and monocyte chemoattractant protein-1 immediately after herniation: an experimental study using a new hernia model. *Spine* 2005;30:55-61.
 134. Kato T, Haro H, Komori H, Shinomiya K. Sequential dynamics of inflammatory cytokine, angiogenesis inducing factor and matrix degrading enzymes during spontaneous resorption of the herniated disc. *J Orthop Res* 2004;22:895-900.
 135. Roberts S, Evans H, Menage J, et al. TNFalpha-stimulated gene product (TSG-6) and its binding protein, Ialpa1, in the human intervertebral disc: new molecules for the disc. *Eur Spine J* 2005;14:36-42.

136. Grang L, Gaudin P, Trocme C, Phelip X, Morel F, Juvin R. Intervertebral disk degeneration and herniation: the role of metalloproteinases and cytokines. *Joint Bone Spine* 2001;68:547-553.
137. Liu J, Roughley PJ, Mort JS. Identification of human intervertebral disc stromelysin and its involvement in matrix degradation. *J Orthop Res* 1991;9:568-575.
138. Matsui Y, Maeda M, Nakagami W, Iwata H. The involvement of matrix metalloproteinases and inflammation in lumbar disc herniation. *Spine* 1998;23:863-868.
139. Kanemoto M, Hukuda S, Komiya Y, Katsuura A, Nishioka J. Immunohistochemical study of matrix metalloproteinase-3 and tissue inhibitor of metalloproteinase-1 human intervertebral discs. *Spine* 1996;21:1-8.
140. Haro H, Crawford HC, Fingleton B, et al. Matrix metalloproteinase-3-dependent generation of a macrophage chemoattractant in a model of herniated disc resorption. *J Clin Invest* 2000;105:133-141.
141. Karelina TV, Goldberg GI, Eisen AZ. Matrix metalloproteinases in blood vessel development in human fetal skin and in cutaneous tumors. *J Invest Dermatol* 1995;105:411-417.
142. Minamide A, Hashizume H, Yoshida M, Kawakami M, Hayashi N, Tamaki T. Effects of basic fibroblast growth factor on spontaneous resorption of herniated intervertebral discs: an experimental study in the rabbit. *Spine* 1999;24:940-945.
143. Haro H, Kato T, Komori H, Osada M, Shinomiya K. Vascular endothelial growth factor (VEGF)-induced angiogenesis in herniated disc resorption. *J Orthop Res* 2002;20:409-415.
144. Franson RC, Saal JS, Saal JA. Human disc phospholipase A2 is inflammatory. *Spine* 1992;17(6 suppl):S129-S132.
145. Saal JS, Franson RC, Dobrow R, Saal JA, White AH, Goldthwaite N. High levels of inflammatory phospholipase A2 activity in lumbar disc herniations. *Spine* 1990;15:674-678.
146. Saal JS. The role of inflammation in lumbar pain. *Spine* 1995;20:1821-1827.
147. Pruzanski W, Vadas P. Phospholipase A2—a mediator between proximal and distal effectors of inflammation. *Immunol Today* 1991;12:143-146.
148. Miyamoto H, Saura R, Doita M, Kurosaka M, Mizuno K. The role of cyclooxygenase-2 in lumbar disc herniation. *Spine* 2002;27:2477-2483.
149. Zelenka M, Schafers M, Sommer C. Intraneural injection of interleukin-1beta and tumor necrosis factor-alpha into rat sciatic nerve at physiological doses induces signs of neuropathic pain. *Pain* 2005;116:257-263.
150. Sox H, Stern S, Owens D, Abrams HL. Assessment of diagnostic technology in health care: rationale, methods, problems and directions. Washington, DC: National Academy Press, 1989.
151. Wiesel SW, Tsourmas N, Feffer HL, Citrin CM, Patronas N. A study of computer-assisted tomography. I. The incidence of positive CAT scans in an asymptomatic group of patients. *Spine* 1984;9:549-551.
152. Boden SD, Davis DO, Dina TS, Patronas NJ, Wiesel SW. Abnormal magnetic-resonance scans of the lumbar spine in asymptomatic subjects: a prospective investigation. *J Bone Joint Surg Am* 1990;72:403-408.
153. Jensen MC, Brant-Zawadzki MN, Obuchowski N, Modic MT, Malkasian D, Ross JS. Magnetic resonance imaging of the lumbar spine in people without back pain. *N Engl J Med* 1994;331:69-73.
154. Saal JA, Saal JS, Herzog RJ. The natural history of lumbar intervertebral disc extrusions treated nonoperatively. *Spine* 1990;15:683-686.
155. Modic MT, Ross JS, Obuchowski NA, Browning KH, Cianflocco AJ, Mazanec DJ. Contrast-enhanced MR imaging in acute lumbar radiculopathy: a pilot study of the natural history. *Radiology* 1995;195:429-435.
156. Modic MT, Obuchowski NA, Ross JS, et al. Acute low back pain and radiculopathy. *Radiology* 2005;237:597-604.
157. Benoist M. The natural history of lumbar degenerative spinal stenosis. *Joint Bone Spine* 2002;69:450-457.
158. Carragee EJ, Kim DH. A prospective analysis of magnetic resonance imaging findings in patients with sciatica and lumbar disc herniation: correlation of outcomes with disc fragment and canal morphology. *Spine* 1997;22:1650-1660.
159. Vital JM, Gille O, Pointillart V, et al. Course of Modic 1 6 months after lumbar posterior osteosynthesis. *Spine* 2003;28:715-720.
160. Weishaupt D, Zanetti M, Hodler J, et al. Painful lumbar disk derangement: relevance of endplate abnormalities at MR imaging. *Radiology* 2001;218:420-427.
161. Braithwaite I, White J, Saifuddin A, Renton P, Taylor BA. Vertebral end-plate (Modic) changes on lumbar spine MRI: correlation with pain reproduction at lumbar discography. *Eur Spine J* 1998;7:363-368.
162. Sandhu HS, Sanchez-Caso LP, Parvataneni HK, Cammisia FP Jr, Girardi FP, Ghelman B. Association between findings of provocative discography and vertebral endplate signal changes as seen on MRI. *J Spinal Disord* 2000;13:438-443.
163. Buttermann GR, Heithoff KB, Ogilvie JW, Transfeldt EE, Cohen M. Vertebral body MRI related to lumbar fusion results. *Eur Spine J* 1997;6:115-120.

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17-20	\$913	\$941	\$1,641	\$2,412	\$3,169	\$3,929
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